Compensatory changes in GABA and NMDA synaptic currents help recover synchronous activity during chronic AMPAergic blockade

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The emergence of synchronous bursts of action potentials is a hallmark of cultured neuronal networks during the first few weeks in vitro [1]. While it has often been assumed that pharmacological blockade of AMPAergic transmission using CNQX eliminates population bursts, we have shown that this effect is transient, and when CNQX is applied chronically, bursting returns within hours. Here, we sought to determine how population bursting homeostatically recovers in the absence of AMPAergic transmission and examined how other transmitter systems contributed to this recovery. Whole-cell voltage clamp recordings revealed an increase in the amplitude of inward synaptic currents within an hour of AMPAergic blockade. MEA recordings showed that NMDA mediates the recovery of population bursting during the first 48 hours of AMPAergic blockade, but that GABA ergic transmission alone was sufficient to recover bursting during a more extended glutamatergic blockade. Together these results suggest that impaired AMPAergic transmission triggers compensatory changes in two other transmitter systems that help recover normal patterns of network activity. This form of plasticity offers a unique strategy for maintaining excitatory drive in cortical circuits when other avenues for excitation are chronically impaired.

Abbreviations: MEA microelectrode array • DIV days in vitro • CNQX 6-cyano-7-nitroquinoxaline-2,3-dione • **APV** D(-)-2-amino-5-phosphonopentanoic acid

Methods

Dissociated cortical cultures

- primary cultures of neurons and glia from E18 rat cortex
- polyethyleneimine- and laminin-coated coverslips or MEAs
- serum-containing growth medium changed every 3 days
- stored in incubator regulated at 35° C and 5% CO
- sterility and humidity maintained with Teflon or polydimethylsiloxane lids [2,3]

Drug treatments

- 40µM CNQX used to block AMPAergic transmission
- 50µM APV used to block NMDAergic transmission
- 20µM bicuculline used to block GABAergic transmission

MEA electrophysiology

- continuously recorded extracellular spikes from cultures grown on MCS 6-well MEAs for several days
- Neurorighter open-source, real-time electrophysiology platform used for multichannel data acquisition [4]
- all recordings performed in standard growth medium inside of incubator

Whole-cell recordings

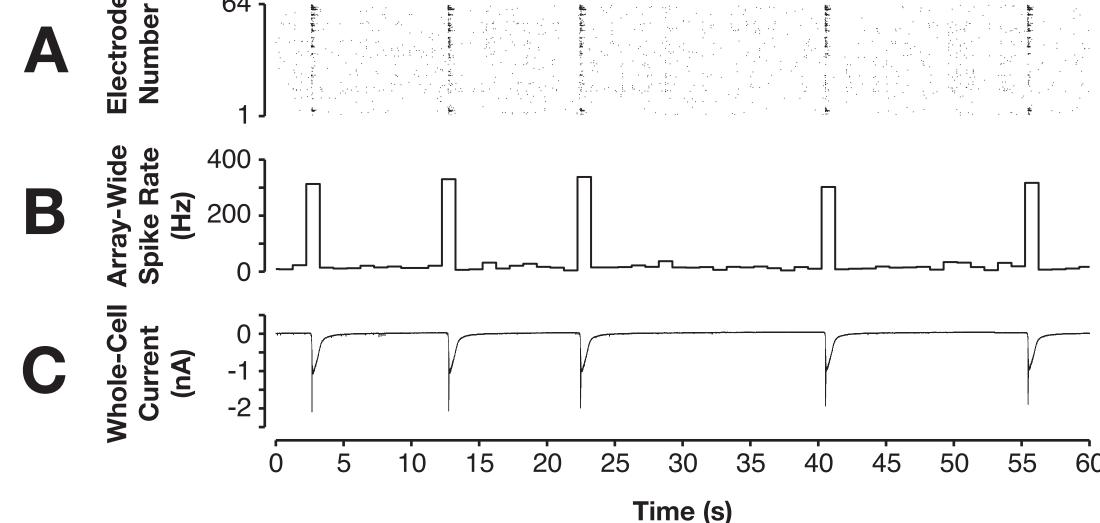
- whole-cell voltage clamp used to record postsynaptic currents from pyramidal shaped cells in cultures grown on coverslips
- HEKA EPC8 amplifier and Patchmaster software used for acquisition
- all recordings performed in continuous perfusion of artificial cerebrospinal fluid bubbled with 5% CO2 and 95% 02

References:

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during population bursts.

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- [4] Rolston JD, Gross R, Potter SM (2009). A low-cost multielectrode system for data acquisition enabling real-time closedloop processing with rapid recover from stimulation artifacts. *Frontiers in Neuroengineering* 2.



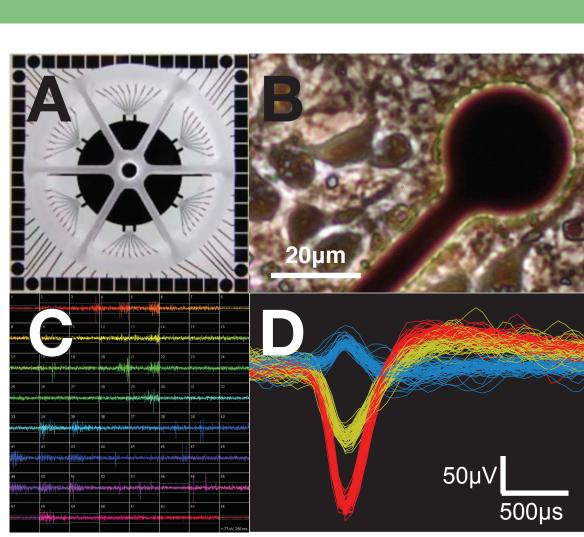


Figure 1: Microelecrode arrays. (A) Multiwell MEA with six pharmacologically-isolated recording chambers. (B) Phase-contrast micrograph of neurons near an MEA electrode. (C) Voltage traces recorded on each electrode using the Neurorighter acquisition system. (D) Sorted spike waveforms recorded on single MEA electrode.

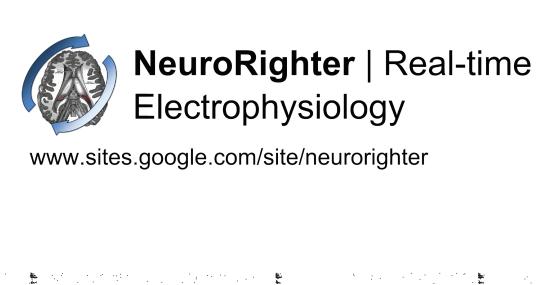


Figure 2: Population bursts can be assessed using MEA or whole-cell recordings. (A) Rastergram showing MEA-recorded spike times by electrode number. Five population bursts are shown. (B) Time histogram of spike times in A, collapsed across entire MEA. Bin size, 1s. (C) Whole-cell recording showing intracellular currents elicited

